



Contents lists available at ScienceDirect

Biochemical and Biophysical Research Communications

journal homepage: www.elsevier.com/locate/ybbrc



Phosphorylation of sterol regulatory element binding protein-1a by protein kinase A (PKA) regulates transcriptional activity



Qingming Dong^a, Francesco Giorgianni^b, Xiong Deng^{a,d}, Sarka Beranova-Giorgianni^b, Dave Bridges^c,
Edwards A. Park^a, Rajendra Raghoe^{a,d,1}, Marshall B. Elam^{a,d,*,1}

^a Department of Pharmacology, University of Tennessee Health Science Center, Memphis, TN, USA

^b Department of Pharmaceutical Sciences, University of Tennessee Health Science Center, Memphis, TN, USA

^c Department of Physiology, University of Tennessee Health Science Center, Memphis, TN, USA

^d Research Service, Veteran's Affairs Medical Center, Memphis, TN, USA

ARTICLE INFO

Article history:

Received 3 May 2014

Available online 20 May 2014

Keywords:

Sterol regulatory element binding protein (SREBP)

Protein kinase A (PKA)

Immobilized metal affinity chromatography (IMAC)

Phosphorylation

Site-directed mutagenesis

Mass spectrometry (MS)

ABSTRACT

The counter-regulatory hormone glucagon inhibits lipogenesis via downregulation of sterol regulatory element binding protein 1 (SREBP-1). The effect of glucagon is mediated via protein kinase A (PKA). To determine if SREBP-1 is a direct phosphorylation target of PKA, we conducted mass spectrometry analysis of recombinant n-terminal SREBP-1a following PKA treatment *in vitro*. This analysis identified serines 331/332 as bona-fide phosphorylation targets of PKA. To determine the functional consequences of phosphorylation at these sites, we constructed mammalian expression vector for both nSREBP-1a and 1c isoforms in which the candidate PKA phosphorylation sites were mutated to active phosphomimetic or non-phosphorylatable amino acids. The transcriptional activity of SREBP was reduced by the phosphomimetic mutation of S332 of nSREBP-1a and the corresponding serine (S308) of nSREBP-1c. This site is a strong candidate for mediating the negative regulatory effect of glucagon on SREBP-1 and lipogenesis.

Published by Elsevier Inc.

1. Introduction

Obesity is a major public health issue in the developed world. The prevalence of obesity is closely correlated with the increased incidence of major diseases such as type 2 diabetes (T2DM), non-alcoholic fatty liver disease, atherosclerotic heart disease, etc. [1,2]. Although the molecular mechanisms by which obesity causes its associated diseases are still unknown, one common feature is dysregulation of lipid homeostasis [1,3]. Among known lipogenic regulators, the sterol regulatory element-binding proteins (SREBPs) are pivotal activators of enzymes responsible for biosynthesis of fatty acids and cholesterol [4,5]. Therefore, understanding the underlying molecular mechanisms that inhibit the SREBPs is important for development of effective treatments of the dyslipidemia that accompanies obesity and T2DM.

In mammals, there are three SREBPs including SREBP-1a, -1c and -2, which are encoded by two genes, *srebf1* and *srebf2* [6]. SREBPs are basic helix loop helix leucine zipper (b/HLH/LZ) transcription factors [7,8]. SREBP-2 controls cholesterol homeostasis

by activating genes mediating cholesterol synthesis and lipoprotein uptake [7]. The SREBP-1 isoforms (1a and 1c) are the predominant inducers of hepatic fatty acid and triglyceride synthesis. The target genes of SREBP-1 include the rate-limiting lipogenic enzymes fatty acid synthase (FASN), stearoyl-CoA desaturase (SCD) and acetyl Co-A carboxylase-1 (ACC-1) [9,10]. Aberrant regulation of SREBP-1c has been implicated in the pathogenesis of dyslipidemia and hepatic steatosis [8,11]. In this regard, understanding the molecular mechanisms by which hormones which suppress lipogenesis such as glucagon inhibit SREBP-1 is critically important.

In the fasting state, glucagon reduces hepatic lipid synthesis [12]. The metabolic effects of glucagon are mediated through the cAMP-dependent protein kinase A (PKA) [13,14]. In animal models, PKA activation by adrenergic stimulation results in lean phenotypes and improves insulin sensitivity [15,16]. PKA activation attenuates SREBP-1 activity and SREBP-1-mediated lipogenesis [16,17]. We have previously shown that glucagon and dibutyryl-cAMP effectively block activation of SREBP-1c transcription by insulin [18,19] and further that db-cAMP inhibits the ER to Golgi transport and proteolytic activation of nascent SREBP-1c [20,21]. However the mechanism(s) underlying the ability of glucagon and its surrogate cAMP to inhibit SREBP-1c remain unclear.

* Corresponding author. Address: Department of Pharmacology, University of Tennessee Health Science Center, 874 Union Ave., Memphis, TN 38163, USA.

E-mail address: melam@uthsc.edu (M.B. Elam).

¹ These authors contributed equally to this manuscript.

A growing body of evidence suggests that posttranslational modification of SREBPs regulate their stability and transcriptional activity [21,22]. Phosphorylation of SREBP-1 by PKA has been proposed as mediating inhibition of its transcriptional activity by cAMP [17]. To determine the effect of PKA mediated phosphorylation of SREBP-1c, we identified serines S331/332 of rat SREBP-1a as phosphorylation targets of PKA. We determined that phosphorylation of S332 of rat SREBP-1a and the corresponding S308 of SREBP-1c inhibits transcriptional activity of SREBP-1. This site corresponds to S338 of human SREBP-1a previously implicated by Lu et al. [17] as a potential phosphorylation site for PKA. Our direct identification of the corresponding site on rat SREBP-1a by mass spectrometry analysis further strengthens the biologic relevance of this site. Based on these findings, it is likely that phosphorylation of this serine by PKA mediates the negative regulatory effect of cAMP on SREBP-1.

2. Materials and methods

2.1. Bacterial expression plasmid construction

Bacterial expression plasmid pET28a-nSREBP-1a (1–403aa), driven by T7 promoter, was constructed by inserting the coding sequence of amino acids (aa) 1–403 of rat SREBP-1a into pET28a vector on the basis of pSREBP-1a (NCBI Reference Sequence: NP_001263636.1). The coding sequence was amplified by PCR from rat hepatocyte cDNA using HotStar HiFidelity Taq Polymerase kit (Qiagen). The forward primer (EcoRI sited added) was CCGGAATTC ATGGACGAGCTACCTTCGGT, and the reverse primer (XhoI sited added) was CCGCTCGAGTGTGCTCCTCCACTGCCACAA. The PCR conditions were 95 °C for 5 min, 35 cycles: 94 °C 15 s, 50 °C 1 min, 68 °C 2 min, last extension at 72 °C for 10 min.

2.2. Mammalian expression plasmid construction

The coding sequence of rat nSREBP-1a was amplified by PCR from rat hepatocyte cDNA using HotStar HiFidelity Taq Polymerase kit. The PCR conditions were as follows. SREBP-1a-F: CCGGAATTC ATGGACGAGCTACCTTCGGT (EcoRI site added) and SREBP-1a-R: CCGCTCGAGCAGGGCCAGGCGGGAGCG (XhoI site added). The insert of nSREBP-1a was cloned into pcDNA3.1/Zeo+ between EcoRI and XhoI sites. The coding sequence of rat nSREBP-1c was amplified by PCR from rat hepatocyte cDNA. The PCR primers were SREBP-1c-EcoRI-F: CCGGAATTCACCATGGATTGCACATTGAAG ACATG (EcoRI site added, Kozak sequence added) and SREBP-1c-XhoI-R: CCGCTCGAGCAGGGCCAGGCGGGAGCG (XhoI site added). Thermal Cycle conditions were described above. The insert of nSREBP-1c was cloned into pcDNA3.1/Zeo+.

2.3. Site-directed mutagenesis

Site-directed mutagenesis was performed according to the manual of QuikChange Site-Directed Mutagenesis Kit from Stratagene. For S308A or S332A, the following primers were used. Mut S to A-F: GAAGCGCTACCGTTCCGCTATCAATGACAAGATTG and Mut S to A-R: CAATCTTGTCATTGATAGCGGAACGGTAGCGCTTC. For S308D or S332D, the following primers were used. Mut S to D-F: GAAGCGCTACCGTTCCGATATCAATGACAAGATTG and Mut S to D-R: CAATCTTGTCATTGATATCGGAACGGTAGCGCTTC.

2.4. Expression of pET28a-nSREBP-1a (1–403aa)

200 ml of LB-broth was inoculated with 5 ml of a stationary phase culture of BL21(DE3) transformed with pET28a-nSREBP-1a (1–403aa) and grown at 37 °C until an absorbance of 0.5 at

590 nm was reached. After adding IPTG to a final concentration of 1 mM, the cells were grown for an additional 4 h at 37 °C.

2.5. Purification of the recombinant nSREBP-1a (1–403aa)

The buffers used in the purification process contained 50 mM NaH₂PO₄, 10 mM Tris, 300 mM NaCl and varying concentrations of imidazole. The lysis buffer contained 50 mM imidazole, the wash buffer 150 mM imidazole and the elution buffer 250 mM imidazole. The transfected BL21(DE3) cells were resuspended in 5 ml of lysis buffer and sonicated with a sonic dismembrator (Fisher Scientific) for 30 s with a 30 s interval 10 times. After centrifugation at 12,000rpm for 15 min at 4 °C, 5 ml of the supernatant was loaded on the 1 ml HisPur Ni-NTA Spin column (Thermo Scientific). Following incubation for 1 h at 4 °C, the resin was washed three times with 5 ml wash buffer. The His-tagged nSREBP-1a was eluted from the resin by adding 0.5 ml elution buffer for four times. The samples were loaded on a SDS–PAGE gel and the gels were stained with GelCode Blue Stain Reagent (Thermo Scientific).

2.6. In vitro PKA phosphorylation

5 µg of purified nSREBP-1a was incubated at 30 °C for 2 h with 2500U PKA under 1× PKA Reaction Buffer (pH 7.5): 50 mM Tris–HCl, 10 mM MgCl₂ and supplemented with/without 200 µM ATP.

2.7. Pro-Q diamond and GelCode Blue staining

Gels were stained with either regular GelCode Blue or the Pro-Q Diamond (Invitrogen). SDS–polyacrylamide gel electrophoresis was performed by standard methods utilizing 5% stacking gels, pH 6.8 and 10% separating gels, pH 8.8. Fluorescent staining of SDS–polyacrylamide gels using Pro-Q Diamond phosphoprotein gel stain was performed by fixing the gels in 45% methanol, 5% acetic acid for 1 h, washing with three changes of deionized water for 10–20 min per wash, followed by incubation in Pro-Q Diamond phosphoprotein gel stain for 90 min, and destaining with 3 washes in 20% acetonitrile in 50 mM sodium acetate, pH 4.0, for 30–60 min each. Before imaging, the gels were rehydrated in deionized water for 30 min. Images were acquired on the stained gels on a 300 nm UV transilluminator from an AlphaImager EP System (Alpha Innotech). GelCode Blue staining was performed according to the manufacture manual and the images were taken using the same system.

2.8. LC–MS/MS

Following SDS–PAGE fractionation, the band corresponding to phosphorylated nSREBP-1a was excised from the gel, the gel pieces placed in a polypropylene tube, de-stained twice with 50% acetonitrile in 50 mM NH₄HCO₃, and dehydrated in a vacuum centrifuge. To digest the phosphoprotein, the gel pieces were re-hydrated for 45 min with a solution containing 100 ng trypsin or Lys-C in 50 mM ammonium bicarbonate. Excess trypsin or Lys-C digestion solution was removed and replaced with 50 mM ammonium bicarbonate; the digestion proceeded for 12 h at 37 °C. The peptide digest solution was collected and residual peptides were extracted from the gel pieces with a solution of 60% acetonitrile/35% water/5% trifluoroacetic acid; the extract was combined with the original digest solution and dried in a vacuum centrifuge. To enrich for phosphopeptides, the digests were subjected to Immobilized Metal Ion Affinity Chromatography (IMAC) as previously described [23]. The enriched digests were analyzed by LC–MS/MS performed with a high-sensitivity system consisting of a nanoflow LC (FAMOS/ULTIMATE from Dionex) coupled to a linear ion trap mass spectrometer (LTQ from ThermoElectron). The LC separation of the

enriched phosphopeptide mixtures was performed on-line with a C18 reversed-phase column, using a fused silica capillary column/spray needle packed in-house with reversed-phase particles (MAGIC C18 from Michrom Bioresources). The peptides eluted from LC were ionized by nano-electrospray and analyzed with the linear ion trap mass spectrometer. The LC was performed at a flow rate of 200 nL/min using a 90-min linear gradient from 0% to 90% mobile phase B. Mobile phase B was 10% water/90% methanol/0.05% formic acid; mobile phase A was 98% water/2% methanol/0.05% formic acid [24]. For each peptide, MS and MS/MS data were acquired in the data-dependent mode. The LC–MS/MS data were used to query the UniProt protein sequence database with the program TurboSEQUEST, part of the Bioworks 3.2 LTQ software. The search parameters included phosphorylation of S, T, Y, and oxidation of M as dynamic modifications, and carbamidomethylation of C as a static modification. The structure-diagnostic fragmentation patterns generated by MS/MS were used to: (a) obtain the primary sequence of the peptide, and (b) locate the phosphorylated amino acid(s) within the peptide.

2.9. Sequence alignment

Human and rat SREBP-1a and 1c protein sequence alignment was performed by Vector NTI 9 and GeneDoc 2.6.

2.10. Immunoblot analysis

Western analysis was conducted as previously described [25]. The immunoblots were scanned and quantified using Quantity One software from Bio-Rad.

2.11. Luciferase assay

HEK293 cells were plated in 48-well plate (2×10^4 cells/well) in DMEM containing 5 mM glucose. On the second day, 250 ng pcDNA3.1 or 220 ng pcDNA3.1-nSREBP-1c (WT, 308A and 308D)/pcDNA3.1-nSREBP-1a (WT, 332A and 332D) plus 30 ng pGL4-FASN-Luciferase (or pGL3-5xSRE-Luciferase) were transfected into the cells. On the fourth day, cells were harvested in 200 μ l passive lysis buffer and luciferase activity was quantified with a fluorometer (Turner Designs) using the Dual-Luciferase Reporter Assay System (Promega).

3. Results

3.1. Expression, purification and *in vitro* PKA phosphorylation of nSREBP-1a

Expression of nSREBP-1a in BL21(DE3) *Escherichia coli* yielded a high level of the recombinant protein shown as a 46 kDa band in total cell extract of induced cells transformed with pET28a-nSREBP-1a (Fig. 1A). Recombinant His-tagged nSREBP-1a was purified with an IMAC column. As shown in Fig. 1B, this purification method yielded highly pure protein fractions.

To confirm that nSREBP-1a was phosphorylated by PKA, we assessed phosphorylation of the protein following incubation with the catalytic subunit PKA-C in the presence and absence of a phosphate donor (ATP) (Fig. 2A). In the presence of ATP, this reaction generated phosphorylated nSREBP-1a, as confirmed with phospho-specific gel staining (Pro-Q Diamond; Fig. 2A). An additional phosphorylated protein band was detected in both ATP-treated and control sample. This band, which appeared at ca. 38 kDa, corresponds to the constitutively phosphorylated catalytic subunit of PKA.

3.2. Mass spectrometry

To identify the phosphorylation sites in nSREBP-1a that are target of PKA, the phosphorylated protein was subjected to digestion followed by LC–MS/MS and database searches. To maximize protein coverage, *in silico* digestion of the protein sequence with different proteases was performed. This *in silico* analysis indicated that Lys-C and trypsin provided complete coverage of potential phosphorylation sites in nSREBP-1a. Thus, these two proteases were selected for the digestion of recombinant phosphorylated nSREBP-1a. To enrich the mixtures for phosphopeptides, the digests were subjected to IMAC purification. The enriched fractions were analyzed by LC–MS/MS, and the datasets used for searches of protein sequence database. A single phosphorylated peptide was revealed. The MS/MS spectrum, shown in Fig. 2B, displays a phosphate-diagnostic product ion corresponding to loss of phosphoric acid from the activated molecular ion. Furthermore, the spectrum contains a number of sequence-determining product ions that match the amino acid sequence of YRSSINDK of rat SREBP-1a. This peptide contains three potential phosphorylation sites. The MS/MS

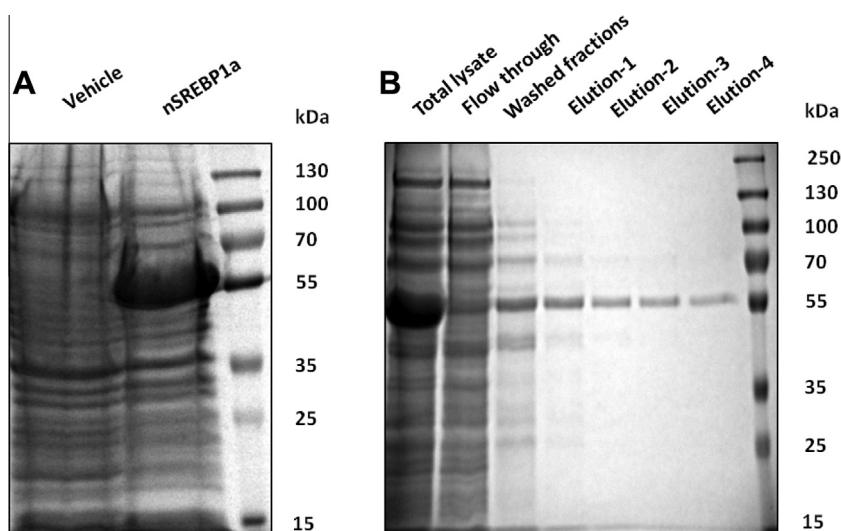


Fig. 1. Expression and purification of nSREBP-1a (1–403aa) in *E. coli*. (A) pET28a and pET28a-nSREBP-1a were transformed into *E. coli* BL21(DE3) and expressed as described in Section 2. The cell extract was loaded on SDS–PAGE gel and analyzed by staining with GelCode Blue Stain Reagent. The two his tagged nSREBP-1a (1–403aa) proteins are about 46 kDa. (B) Eluted fractions from the Ni-NTA resin are shown.

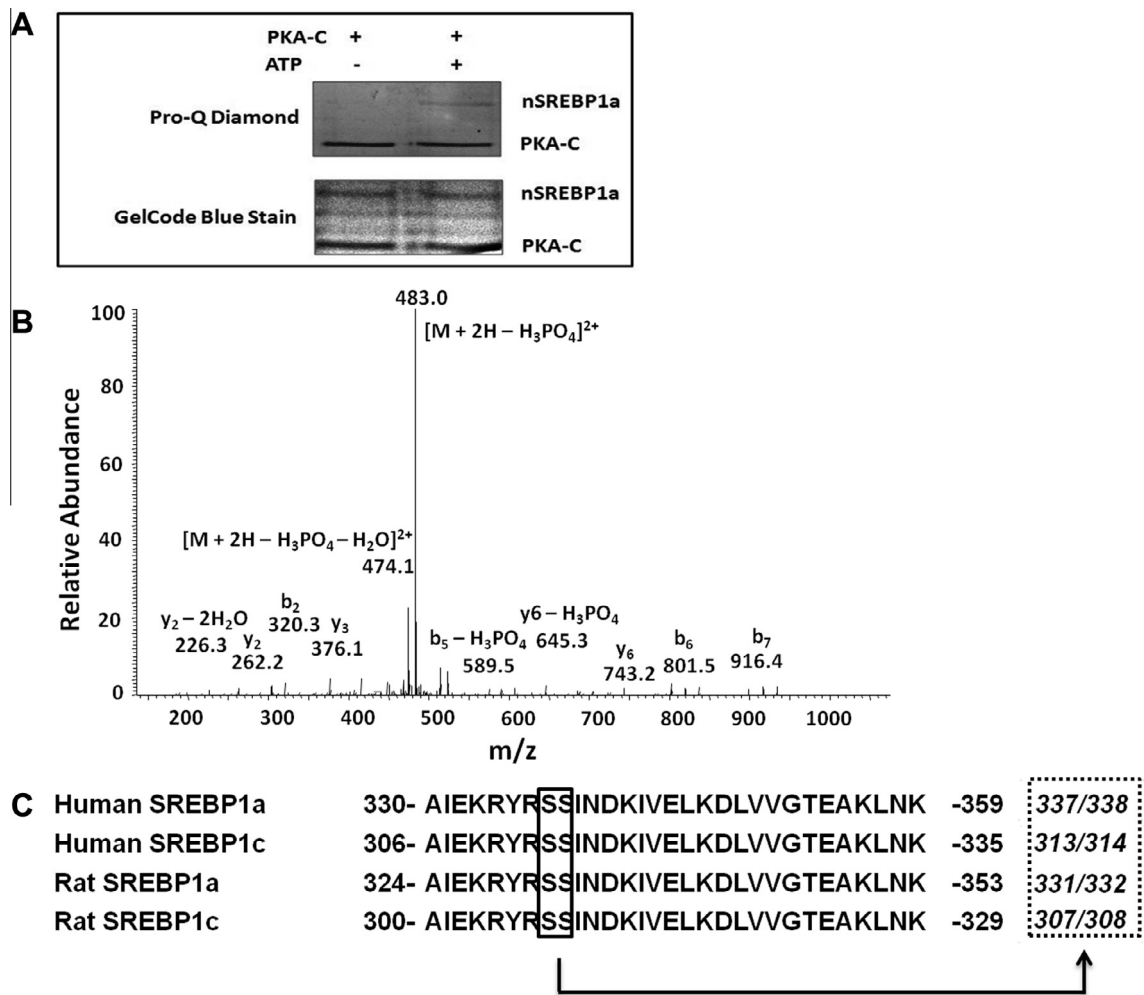


Fig. 2. Mass spectrometry analysis of SREBP-1a. (A) *In vitro* phosphorylation of bacterial expressed nSREBP-1a (1–403aa). Purified nSREBP-1a was treated with PKA-C. The protein was loaded on SDS–PAGE gel and stained with Pro-Q Diamond Phosphoprotein gel stain (upper panel) or with GelCode Blue Stain Reagent (lower panel). The upper band of the GelCode Blue Stain reagent stained bands was cut for LC–MS/MS. (B) Mass spectrometry (LC–MS/MS) analysis of PKA-treated SREBP-1a. The MS/MS spectrum of a phosphorylated peptide that matched the amino acid sequence YRSSINDK of rat SREBP-1a. The fragment ions in the spectrum localize the phosphorylation site at either S331 or S332. (C) Sequence alignment of human and rat SREBP-1a and 1c around the PKA phosphorylation site. The conserved amino acid sequence between human and rat SREBP-1a and 1c is shown.

data (Fig. 2B) exclude Y329 as a phosphorylation candidate site, and localize the phosphorylation to either S331 or S332.

3.3. Sequence alignment

The sequence alignment of human and rat SREBP-1a and 1c in the sequence region around the PKA phosphorylation sites is shown in Fig. 2C. This region is highly conserved between human and rat SREBP-1a and 1c suggesting that PKA phosphorylates human and rat SREBP-1a and 1c *in vivo*.

3.4. Stability and transcriptional activity

We assessed the effect of loss of function (Ser to Ala, S to A) and gain of function (Ser to Asp, S to D) mutations on stability of SREBP1c and 1a. The results showed that the stability of both nSREBP1a and nSREBP1c containing aspartic acid substitution at S332 or S308 (D mutant) was not decreased but rather was slightly increased (Fig. 3A). On the contrary, alanine substitution at S332 or S308 (A mutant) had no effect on stability of nSREBP1a and 1c (Fig. 3A). When the transfected cells were treated with 10 μ M proteasomal inhibitor MG132, the protein levels of WT and both mutants were the same (Fig. 3A). These results showed that phosphomimetic mutation of

the putative PKA sites S332D of nSREBP1a and S308D of nSREBP1c does not decrease SREBP-1a and 1c stability, respectively.

The ability of WT and mutant forms of nSREBP-1 to activate transcription of the known target gene FASN and a synthetic Sterol Response Element (5xSRE) was assessed using luciferase (Luc) assays. In these reporter assays, the level of PKA mutant proteins was normalized to that of WT nSREBP-1a or 1c. WT nSREBP-1a increased relative luciferase activity of pGL4-FASN-Luc and 5xSRE about 35 and 5.8-fold, respectively, as compared to mock vector transfected cells (Fig. 3B). Similarly WT nSREBP-1c increased relative luciferase activity of pGL4-FASN-Luc and 5xSRE about 31 and 3.9-fold, respectively (Fig. 3B). Mutation of S332 and S308 to the non-phosphorylatable amino acid alanine (332A, 308A) of n-terminal SREBP-1a and 1c enhanced transactivation of the luciferase report genes FASN and 5xSRE (Fig. 3B). Conversely, mutation of these serines to the phosphomimetic amino acid aspartic acid (332D, 308D) strongly inhibited the transactivating capacity of nSREBP-1a and 1c (Fig. 3B).

4. Discussion

During fasting or acute stress, the biosynthesis of lipids in the liver is inhibited in part by cAMP [26]. Glucagon, adrenaline and

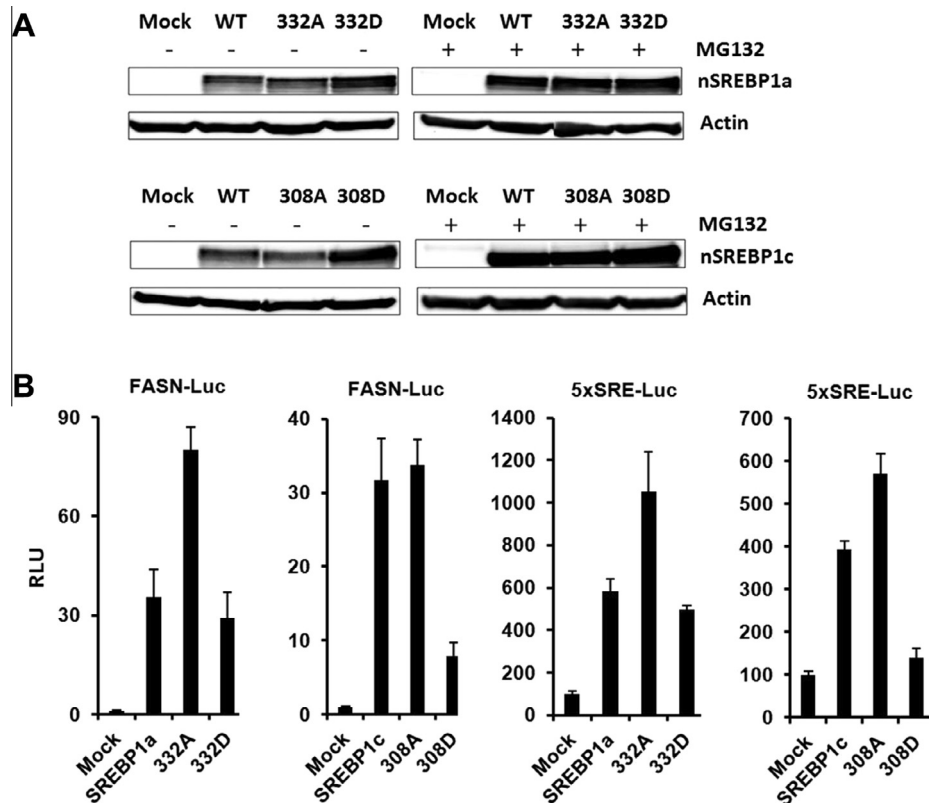


Fig. 3. Stability and transcriptional activity of SREBP mutants. (A) HEK293 cells were transfected with expression plasmids encoding nSREBP-1a (WT, 332A and 332D) or nSREBP-1c (WT, 308A and 308D). After incubation for 18 h, the cells were treated with/without proteasomal inhibitor MG132 (10 μ M) for 6 h. Total cell lysates were extracted and Western blots were performed. Actin was used as the loading control. (B) HEK293 cells were cotransfected with a luciferase reporter (Luc) containing the promoter region of the FASN gene (FASN-Luc) or 5xSRE-Luc along with expression plasmids encoding either the WT or the mutant forms of nSREBP-1a or nSREBP-1c. Luciferase assays were conducted. Data represent the means \pm SEMs of three independent experiments. During reporter assays, the level of mutant proteins was normalized to that of WT nSREBP-1a or 1c.

other hormones that raise cAMP levels reduce the activity and abundance of lipogenic enzymes, which are critical for the synthesis of fatty acids and triglycerides [27,28]. Glucagon represses SREBP-1c activity through PKA signaling cascades [17,20]. In addition, the expression of SREBP-1c is suppressed by fasting or nutritional deprivation [29,30]. Although the activity of SREBP-1c is regulated by PKA or AMP-activated protein kinases (AMPKs), the exact underlying mechanisms have yet to be determined *in vivo* [17,31].

As the key regulatory factor in lipogenesis, SREBPs are targets of hormones such as insulin, glucagon, and growth factors [19,32,33]. Activation of SREBPs is tightly regulated at multiple levels, including transcription, precursor maturation, nuclear protein stability and transcriptional activity. The abundance of the nuclear form of SREBPs is controlled by transcriptional upregulation followed by proteolytic cleavage. However, increasing evidence supports the hypothesis that posttranslational modifications of SREBPs modulate their transcriptional activity and stability [30,34,35]. Lu et al. [17] identified two PKA consensus sites in the NH2 terminus of human SREBP-1a, S337 and S338 and postulated that S338 might mediate the inhibitory effect of PKA activation on SREBP-1a. Using unbiased mass spectrometry, we have directly confirmed phosphorylation of the corresponding serines (S331/S332) of rat SREBP-1a by PKA. Furthermore, we determined that phosphorylation of S332 and the corresponding site (S308) of rat SREBP-1c reduces transcriptional activity. Thus it is likely that phosphorylation of SREBP-1 isoforms at this site mediates the inhibitory effect of cAMP on SREBP-1. Recently, Lee et al. [36] linked phosphorylation of SREBP-1c to sumoylation at Lys98 leading to enhanced ubiquitination and degradation.

They observed that the extent of sumoylation and ubiquitination was reduced following mutation of S308 to alanine in nSREBP-1c and postulated that sumoylation may be regulated by phosphorylation of this serine by PKA [36]. Interestingly, in our studies we observed that mutation of this site, either in the context of full-length nascent SREBP-1c or its n-terminal fragment nSREBP-1c did not appreciably alter steady state levels of SREBP-1c protein. This does not eliminate a role of PKA mediated phosphorylation in stability of SREBP-1c protein but does not support the hypothesis that phosphorylation of this serine mediates the effect of PKA on stability. Rather, our data indicate that the predominant effect of PKA mediated phosphorylation SREBP1a at S332 and the corresponding site on SREBP-1c (S308) is to decrease its transcriptional activity.

In conclusion, we used a discovery based mass spectrometry approach to find the phosphorylation target sites of PKA in SREBP-1a. With this unbiased methodology, we have identified S331/S332 of SREBP-1a as a phosphorylation target of PKA. Mutational analysis of this site confirmed that phosphorylation of SREBP-1a at S332 and SREBP-1c at S308 by PKA decreases transcriptional activity of nSREBP-1a and 1c. This site is a strong candidate for mediating the negative regulatory effect of glucagon on SREBP-1c and lipogenesis.

Acknowledgments

This project was funded by VA Research and Development Program Merit Review Grants to M.B.E. and R.R. Senior Research Career Scientist of the Department of Veteran's affairs Funds for the LTQ mass spectrometer were provided by the NIH Shared

Instrumentation Grant S10RR16679, and by the UTHSC College of Pharmacy.

References

- [1] G.M. Reaven, Insulin resistance: the link between obesity and cardiovascular disease, *Med. Clin. North Am.* 95 (2011) 875–892.
- [2] O. Kaidar-Person, G. Bar-Sela, B. Person, The two major epidemics of the twenty-first century: obesity and cancer, *Obes. Surg.* 21 (2011) 1792–1797.
- [3] O. Cheung, A.J. Sanyal, Abnormalities of lipid metabolism in nonalcoholic fatty liver disease, *Semin. Liver Dis.* 28 (2008) 351–359.
- [4] P. Ferre, F. Foufelle, SREBP-1c transcription factor and lipid homeostasis: clinical perspective, *Horm. Res.* 68 (2007) 72–82.
- [5] T.I. Jeon, T.F. Osborne, SREBPs: metabolic integrators in physiology and metabolism, *Trends Endocrinol. Metab.* 23 (2012) 65–72.
- [6] Z. Xiaoping, Y. Fajun, Regulation of SREBP-mediated gene expression, *Sheng Wu Wu Li Hsueh Bao* 28 (2012) 287–294.
- [7] J.D. Horton, J.L. Goldstein, M.S. Brown, SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver, *J. Clin. Invest.* 109 (2002) 1125–1131.
- [8] R. Raghoe, X. Yellaturu, X. Deng, E.A. Park, M.B. Elam, SREBPs: the crossroads of physiological and pathological lipid homeostasis, *Trends Endocrinol. Metab.* 19 (2008) 65–73.
- [9] T.F. Osborne, P.J. Espenshade, Evolutionary conservation and adaptation in the mechanism that regulates SREBP action: what a long, strange trip it's been, *Genes Dev.* 23 (2009) 2578–2591.
- [10] D. Eberle, B. Hegarty, P. Bossard, P. Ferre, F. Foufelle, SREBP transcription factors: master regulators of lipid homeostasis, *Biochimie* 86 (2004) 839–848.
- [11] J.D. Browning, J.D. Horton, Molecular mediators of hepatic steatosis and liver injury, *J. Clin. Invest.* 114 (2004) 147–152.
- [12] E.R. Kandel, The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CPEB, *Mol. Brain* 5 (2012) 14.
- [13] T.O. Chan, J.M. Pascal, R.S. Armen, U. Rodeck, Autoregulation of kinase dephosphorylation by ATP binding in AGC protein kinases, *Cell Cycle* 11 (2012) 475–478.
- [14] G.S. McKnight, D.E. Cummings, P.S. Amieux, M.A. Sikorski, E.P. Brandon, J.V. Planas, K. Motamed, R.L. Idzerda, Cyclic AMP, PKA, and the physiological regulation of adiposity, *Recent Prog. Horm. Res.* 53 (1998) 139–159 (discussion 160–161).
- [15] D.E. Cummings, E.P. Brandon, J.V. Planas, K. Motamed, R.L. Idzerda, G.S. McKnight, Genetically lean mice result from targeted disruption of the RII beta subunit of protein kinase A, *Nature* 382 (1996) 622–626.
- [16] S.A. Schreyer, D.E. Cummings, G.S. McKnight, R.C. LeBoeuf, Mutation of the RIIbeta subunit of protein kinase A prevents diet-induced insulin resistance and dyslipidemia in mice, *Diabetes* 50 (2001) 2555–2562.
- [17] M. Lu, J.Y. Shyy, Sterol regulatory element-binding protein 1 is negatively modulated by PKA phosphorylation, *Am. J. Physiol. Cell Physiol.* 290 (2006) C1477–C1486.
- [18] X. Deng, L.M. Cagen, H.G. Wilcox, E.A. Park, R. Raghoe, M.B. Elam, Regulation of the rat SREBP-1c promoter in primary rat hepatocytes, *Biochem. Biophys. Res. Commun.* 290 (2002) 256–262.
- [19] M. Foretz, C. Pacot, I. Dugail, P. Lemarchand, C. Guichard, X. Le Liepvre, C. Berthelie-Lubrano, B. Spiegelman, J.B. Kim, P. Ferre, F. Foufelle, ADD1/SREBP-1c is required in the activation of hepatic lipogenic gene expression by glucose, *Mol. Cell. Biol.* 19 (1999) 3760–3768.
- [20] C.R. Yellaturu, X. Deng, L.M. Cagen, H.G. Wilcox, E.A. Park, R. Raghoe, M.B. Elam, Posttranslational processing of SREBP-1 in rat hepatocytes is regulated by insulin and cAMP, *Biochem. Biophys. Res. Commun.* 332 (2005) 174–180.
- [21] C.R. Yellaturu, X. Deng, E.A. Park, R. Raghoe, M.B. Elam, Insulin enhances the biogenesis of nuclear sterol regulatory element-binding protein (SREBP)-1c by posttranscriptional down-regulation of Insig-2A and its dissociation from SREBP cleavage-activating protein (SCAP). SREBP-1c complex, *J. Biol. Chem.* 284 (2009) 31726–31734.
- [22] J. Kotzka, B. Knebel, J. Haas, L. Kremer, S. Jacob, S. Hartwig, U. Nitzgen, D. Muller-Wieland, Preventing phosphorylation of sterol regulatory element-binding protein 1a by MAP-kinases protects mice from fatty liver and visceral obesity, *PLoS One* 7 (2012) e32609.
- [23] L. Chen, B. Fang, F. Giorgianni, J.R. Gingrich, S. Beranova-Giorgianni, Investigation of phosphoprotein signatures of archived prostate cancer tissue specimens via proteomic analysis, *Electrophoresis* 32 (2011) 1984–1991.
- [24] F. Giorgianni, A. Cappiello, S. Beranova-Giorgianni, P. Palma, H. Trufelli, D.M. Desiderio, LC-MS/MS analysis of peptides with methanol as organic modifier: improved limits of detection, *Anal. Chem.* 76 (2004) 7028–7038.
- [25] C.R. Yellaturu, X. Deng, L.M. Cagen, H.G. Wilcox, C.M. Mansbach 2nd, S.A. Siddiqi, E.A. Park, R. Raghoe, M.B. Elam, Insulin enhances post-translational processing of nascent SREBP-1c by promoting its phosphorylation and association with COPII vesicles, *J. Biol. Chem.* 284 (2009) 7518–7532.
- [26] D.M. Tarlow, P.A. Watkins, R.E. Reed, R.S. Miller, E.E. Zwerger, M.D. Lane, Lipogenesis and the synthesis and secretion of very low density lipoprotein by avian liver cells in nonproliferating monolayer culture. Hormonal effects, *J. Cell Biol.* 73 (1977) 332–353.
- [27] J. Girard, D. Perdureau, F. Foufelle, C. Prip-Buus, P. Ferre, Regulation of lipogenic enzyme gene expression by nutrients and hormones, *FASEB J.* 8 (1994) 36–42.
- [28] P. Lefevre, C. Diot, P. Legrand, M. Douaire, Hormonal regulation of stearoyl coenzyme-A desaturase 1 activity and gene expression in primary cultures of chicken hepatocytes, *Arch. Biochem. Biophys.* 368 (1999) 329–337.
- [29] J.D. Horton, Y. Bashmakov, I. Shimomura, H. Shimano, Regulation of sterol regulatory element binding proteins in livers of fasted and refed mice, *Proc. Natl. Acad. Sci. U.S.A.* 95 (1998) 5987–5992.
- [30] J.B. Kim, P. Sarraf, M. Wright, K.M. Yao, E. Mueller, G. Solanes, B.B. Lowell, B.M. Spiegelman, Nutritional and insulin regulation of fatty acid synthetase and leptin gene expression through ADD1/SREBP1, *J. Clin. Invest.* 101 (1998) 1–9.
- [31] G. Zhou, R. Myers, Y. Li, Y. Chen, X. Shen, J. Fenyk-Melody, M. Wu, J. Ventre, T. Doebber, N. Fujii, N. Musi, M.F. Hirshman, L.J. Goodyear, D.E. Moller, Role of AMP-activated protein kinase in mechanism of metformin action, *J. Clin. Invest.* 108 (2001) 1167–1174.
- [32] R.J. Mason, T. Pan, K.E. Edeen, L.D. Nielsen, F. Zhang, M. Longphre, M.R. Eckart, S. Neben, Keratinocyte growth factor and the transcription factors C/EBP alpha, C/EBP delta, and SREBP-1c regulate fatty acid synthesis in alveolar type II cells, *J. Clin. Invest.* 112 (2003) 244–255.
- [33] I. Shimomura, Y. Bashmakov, S. Ikemoto, J.D. Horton, M.S. Brown, J.L. Goldstein, Insulin selectively increases SREBP-1c mRNA in the livers of rats with streptozotocin-induced diabetes, *Proc. Natl. Acad. Sci. U.S.A.* 96 (1999) 13656–13661.
- [34] G. Roth, J. Kotzka, L. Kremer, S. Lehr, C. Lohaus, H.E. Meyer, W. Krone, D. Muller-Wieland, MAP kinases Erk1/2 phosphorylate sterol regulatory element-binding protein (SREBP)-1a at serine 117 in vitro, *J. Biol. Chem.* 275 (2000) 33302–33307.
- [35] A. Sundqvist, M.T. Bengoechea-Alonso, X. Ye, V. Lukiyanichuk, J. Jin, J.W. Harper, J. Ericsson, Control of lipid metabolism by phosphorylation-dependent degradation of the SREBP family of transcription factors by SCF(Fbw7), *Cell Metab.* 1 (2005) 379–391.
- [36] G.Y. Lee, H. Jang, J.H. Lee, J.Y. Huh, S. Choi, J. Chung, J.B. Kim, PIASy-mediated sumoylation of SREBP1c regulates hepatic lipid metabolism upon fasting signaling, *Mol. Cell. Biol.* 34 (2014) 926–938.